

AC/2024/RS 2



College of Arts,  
Science &  
Commerce

RISE WITH EDUCATION

(Autonomous) Sion (West), Mumbai – 400022

**Faculty: Science**

**Program: B.Sc.**

**Subject: MICROBIOLOGY**

**Academic Year: 2024– 2025**

**AS PER NATIONAL EDUCATION POLICY 2020**

**Choice Based Credit System (CBCS)**

**S.Y.B.Sc.**

**Revised Credit Based Semester and Grading Syllabus approved by  
Board of Studies in Microbiology brought into effect from June 2024**

## **PROGRAM OUTCOME (PO)**

At the end of an Undergraduate Program, a student would have obtained the following:

- **PO1. Solving Complex Problem:**

Applying the knowledge of various courses learned under a program with an ability to break down complex problems into simple components, by designing processes required for problem solving.

- **PO2. Critical Thinking:**

Organizing thoughts to identify assumptions, verifying the accuracy and validity of assumptions, making informed decisions that guide actions (at Institutional, Personal and Intellectual level), developing the ability to think with different perspectives and ideas.

- **PO3. Reasoning ability and rational thinking:**

Developing rational thinking on the basis of acquired contextual knowledge, assessing societal, public health and safety, cultural, legal, gender, ethnic and environmental issues, and performing with decisive responsibility.

- **PO4. Research skill:**

Utilizing the contextual knowledge in an interdisciplinary framework. Integrating research-based knowledge and research methods involving problem definition, analysis and interpretation of data, synthesis of the information to provide valid conclusions. Exercising analytical skill, research ability, creativity, for employability and collaborating with industries.

- **PO5. Effective Communication skill:**

Facilitating to speak, read, write and listen effectively through both formal language and in one's own mother tongue, in order to make meaning of the world around. Enabling to comprehend and write effective reports and documentation, make successful presentations, give and receive clear instructions.

- **PO6. Proficiency with ICT:**

Equipping to create, select, apply appropriate tools and techniques, resources through electronic media for the purpose of gathering, analyzing data and drawing inference with an understanding of its merits and demerits.

- **PO7. Social Interactive Skills and team work:**

Eliciting networking with people, mediate disagreement and help reach conclusions in group settings. Functioning effectively as an individual, and as a member in diverse groups, and in multidisciplinary settings exhibiting adaptability, leadership quality and team-building

- **PO8. Ethical values:**

Recognizing and respecting different value systems including one's own, to understand the moral dimensions of one's decisions, intention to help the society and feeling good about it, commitment to professional duties and responsibilities.

- **PO9. Self-directed Learning:**

Acquiring the ability to explore and gain knowledge in independent ways, keep evolving lifelong in the broad context of socio-technological changes.

- **PO10. Sensitization towards Environment and Sustainability:**

Understanding the need for sustainable development and concern for environmental issues, realizing the importance of cohabitation, co-evolution in our achievements of sustainable development goal

- **PO11. Gender Sensitization:**

Demonstrating knowledge and understanding of gender equity-issues and gender justice.

- **PO12. Civic Values and Global Citizenship:**

Expressing empathetic social concern while helping others when their rights are violated, no matter where in the world they live, to act with an informed awareness on issues, to participate in civic life by volunteering for social justice.

### PROGRAMME SPECIFIC OUTCOMES (PSO) (FOR MICROBIOLOGY)

- PSO1: Students will be introduced to the subject of Microbiology which is not taught at the junior college
- PSO2: Eloquence in specific phraseology pertaining to the subject of microbiology.
- PSO3: Familiarize with the theories and techniques of the various areas in microbiology.
- PSO4: Obtain expertise in essential practical techniques required in microbiological analysis and prepare for advance studies.
- PSO5: Discuss the applications of microorganisms in the various fields of microbiology.

Course code	MAJOR PAPER I	
	SEMESTER III	
SIUMIMJ211	<b>Nucleic acid chemistry , Nanotechnology and Analytical techniques</b>	<b>3 Credits (45 lectures)</b>
<b>Unit-I</b>	Nucleic acid chemistry	15 lectures
<b>Unit-II</b>	Nanobiotechnology, biofilms & biosensors with applications	15 lectures
<b>Unit-III</b>	Analytical techniques – I	15 lectures
	<b>PRACTICALS</b>	<b>1 Credit</b>

Course code	MAJOR PAPER II	
	SEMESTER III	
SIUMIMJ212	<b>Water Microbiology , Epidemiology &amp; Immunology</b>	<b>3 Credits (45 lectures)</b>
<b>Unit-I</b>	Potable water & Sewage Microbiology	15 lectures
<b>Unit-II</b>	Epidemiology & Diagnostic Microbiology	15 lectures
<b>Unit-III</b>	Introduction to Immunology	15 lectures
	<b>PRACTICALS</b>	<b>1 Credit</b>

<b>MINOR PAPER I</b>		
<b>Course code</b>	<b>SEMESTER III</b>	
<b>SIUMIMN211</b>	<b>Environmental Microbiology</b>	<b>3 Credits (45 lectures)</b>
<b>Unit-I</b>	Air Microbiology	15 lectures
<b>Unit-II</b>	Soil Microbiology	15 lectures
<b>Unit-III</b>	Fresh water & Marine Microbiology	15 lectures
	<b>PRACTICALS</b>	<b>1 Credit</b>

<b>VOCATIONAL SKILL COURSE</b>		
<b>Course code</b>	<b>SEMESTER III</b>	
<b>SIUMIVS211</b>	<b>Techniques in Microbiology I (Practicals)</b>	<b>2 Credits</b>
	<b>OPEN ELECTIVE (Interdisciplinary)</b>	
	<b>SEMESTER III</b>	
<b>SIUMIOE211</b>	<b>Microbiology in everyday life - I</b>	<b>2 Credits</b>
<b>Unit I</b>	Introduction to biology, food microbiology and immunity	15 lectures
<b>Unit II</b>	Tutorial	15 lectures

<b>Course code</b>	<b>MAJOR PAPER I</b>	
	<b>SEMESTER IV</b>	
<b>SIUMIMJ221</b>	<b>Metabolism and Basic Analytical Techniques</b>	<b>3 Credits (45 lectures)</b>
<b>Unit-I</b>	Introduction To Metabolism and Bioenergetics	15 lectures
<b>Unit-II</b>	Enzyme Kinetics	15 lectures
<b>Unit-III</b>	Analytical techniques II	15 lectures
	<b>PRACTICALS</b>	<b>1 Credit</b>

Course code	MAJOR PAPER II	
	SEMESTER IV	
SIUMIMJ222	Dairy Microbiology, Food Microbiology and Taxonomy	3 Credits (45 lectures)
Unit-I	Dairy Microbiology	15 lectures
Unit-II	Food Microbiology	15 lectures
Unit-III	Microbial Taxonomy	15 lectures
	PRACTICALS	1 Credit

MINOR PAPER II		
Course code	SEMESTER IV	
SIUMIMN221	Industrial Microbiology, Microbial Diversity and Applications Of Microbiology	3 Credits (45 lectures)
Unit-I	Industrial Microbiology	
Unit-II	Microbial diversity in extreme environments and Metagenomics	
Unit-III	Biofertiliser, Biopesticide , Bioremediation	
	PRACTICALS	1 Credit

SKILL ENHANCEMENT COURSE		
Course code	SEMESTER IV	
SIUMISE221	Techniques in Microbiology II (Practicals)	2 Credits
OPEN ELECTIVE (Interdisciplinary)		
SEMESTER IV		
SIUMIOE221	Microbiology in Everyday life - II	2 Credits
Unit I	Introduction to agriculture microbiology, microbial infections and antibiotics	15 lectures
Unit II	Tutorial	15 lectures

<b>Course code</b>	<b>Title</b>	<b>Credits</b>
<b>Semester III</b>		
SIUMIMJ211	MAJOR PAPER I	3
SIUMIMJ212	MAJOR PAPER II	3
SIUMIMN211	MINOR PAPER I	3
SIUMIMJP211	MAJOR PRACTICAL I	1
SIUMIMJP212	MAJOR PRACTICAL II	1
SIUMIMNP211	MINOR PAPER PRACTICAL I	1
SIUMIVS211	VOCATIONAL SKILL COURSE	2
SIUMIOE211	OPEN ELECTIVE COURSE	2
		<b>Total = 16</b>
<b>Semester IV</b>		
SIUMIMJ221	MAJOR PAPER I	3
SIUMIMJ222	MAJOR PAPER II	3
SIUMIMN221	MINOR PAPER II	3
SIUMIMJP221	MAJOR PRACTICAL I	1
SIUMIMJP222	MAJOR PRACTICAL II	1
SIUMIMNP221	MINOR PAPER PRACTICAL II	1
SIUMISE221	SKILL ENHANCEMENT COURSE	2
SIUMIOE221	OPEN ELECTIVE COURSE	2
		<b>Total = 16</b>

## S.Y.B.Sc. MICROBIOLOGY SYLLABUS

### SEMESTER III MAJOR PAPER I

**Course: Nucleic acid chemistry, Nanotechnology and Analytical techniques**

**Course code: SIUMIMJ211**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Understanding the nucleic acid molecules & central dogma, genetic code as the basis of life, DNA mutations and studying their effects on the genetic code and defining the different genetic elements
2. Discriminate between the techniques of nanoparticles, biofilm and biosensor and comparatively evaluate their applications.
3. Understanding the principle and working of spectroscopic and electrophoretic techniques and their applications.

<b>Course code SIUMIMJ211</b>	<b>MAJOR PAPER I Nucleic acid chemistry, Nanotechnology and Analytical techniques</b>	<b>3 CREDITS</b>
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	<b>Nucleic acid chemistry and Genetic elements</b> 1.1 Nucleic acid chemistry- Denaturation of double helical DNA and RNA Nucleic acid from different species can form Hybrids, Nucleotides and nucleic acids undergo non enzymatic transformations, DNA methylation 1.2 Other Functions of nucleotides 1.3 Central dogma of life, Genetic code 1.4 Plasmids and types of Plasmids 1.5 Transposons (Structure and Types)	<b>15L</b>

<p><b>Unit II</b></p>	<p><b>Nanobiotechnology, Biofilms and biosensors with applications</b></p> <p>2.1 Nanobiotechnology:</p> <ul style="list-style-type: none"> <li>• Introduction of Nanobiotechnology,</li> <li>• Types of nanomaterials, nanoparticles, nanocapsules, nanotubes, liposomes, nanogels, Dendrimers, Gold nanoparticles(Definitions)</li> <li>• UV characterization of nanoparticles Application in drug and gene delivery</li> </ul> <p>2.2 Biofilms and biosensors</p> <ul style="list-style-type: none"> <li>• Biofilms: Introduction of biofilms, Types of biofilms, Mechanism of formation of biofilms and applications of biofilms.</li> <li>• Biosensors: Introduction, design, working and applications of biosensors</li> </ul>	<p><b>15L</b></p>
<p><b>Unit III</b></p>	<p><b>Analytical Techniques I</b></p> <p>3.1 Spectroscopic Techniques- Visible, UV and IR spectrophotometry Principles, instrumentation and applications</p> <p>3.2 pH meter: principle, instrumentation and Application</p> <p>3.3 Electrophoresis - General principles, Factors affecting electrophoresis, apparatus, support media- agarose gels, polyacrylamide gels &amp; applications</p>	<p><b>15L</b></p>

## S.Y.B.Sc. MICROBIOLOGY SYLLABUS

### SEMESTER III MAJOR PAPER II

**Course: Water Microbiology, Epidemiology & Immunology**

**Course code: SIUMIMJ212**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Review of freshwater and sewage microbiology and understanding the methods of potability testing and sewage treatment.
2. Study the epidemiological methods of disease and evaluate different diagnostic techniques in diagnostic microbiology.
3. Distinguish between different types of immunity and review their role in disease Control.

<b>Course code SIUMIMJ212</b>	<b>MAJOR PAPER II Water Microbiology, Epidemiology &amp; Immunology</b>	<b>3 CREDITS</b>
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	<b>Water Microbiology-Fresh Water and Sewage Microbiology</b> 1.1 Fresh water environments and micro-organisms found in Springs, rivers and streams, Lakes,marshes and bogs 1.2 Potable water: Definition, water purification, water quality standards and pathogens transmittedthrough water 1.3 Microbiological analysis of water: Indicator organisms and their detection in water-Total Coliforms, Faecal Coliforms and <i>E. coli</i> , Faecal <i>Streptococci</i> , <i>Clostridium perfringens</i> 1.4 Modern Waste Water treatment: Primary,Secondary & Tertiary treatment 1.5 The nature of wastewater and Monitoring of waste water treatment process (BOD,COD) 1.6 Removal of Pathogens by Sewage treatment	<b>15L</b>

	<p>Processes</p> <p>1.7 Oxidation Ponds and Septic tanks</p> <p>1.8 Sludge Processing</p> <p>1.9 Disposal of treated waste water and biosolids.</p>	
<b>Unit II</b>	<p><b>Epidemiology and Diagnostic Microbiology</b></p> <p>2.1 Epidemiological terminology: Epidemiology, sporadic diseases, endemic diseases, Hyperendemic Diseases, Epidemic Diseases, Index Case, Pandemic Disease, Outbreak</p> <p>2.2 The Spread of Infection:</p> <p>a) Reservoirs of infection-Human reservoir, Animal reservoir, non-living reservoir.</p> <p>Transmission of Disease- Contact transmission, Vehicle Transmission and vectors</p> <p>2.3 Nosocomial infection: (1L)</p> <p>Microorganisms in hospitals, compromised host, Transmission and control of Nosocomial infection</p> <p>2.4 Isolation of Pathogens from clinical specimens:</p> <p>a) Growth media and Culture</p> <p>b) Collection of specimens, handling and transport</p> <p>c) Types of specimens and their culture ---Blood, Urine, Faeces, sputum, Cerebrospinal fluid, pus, genital and culture of Anaerobes.</p> <p>2.5 Identification of microorganisms from specimens:</p> <p>a) Microscopy</p> <p>Growth-Dependent Identification Methods</p> <p>2.6 Rapid Methods of Identification: Molecular Methods and Analysis of Metabolic Products:</p> <p>a) Nucleic Acid –Based Detection Methods</p> <p>b) Gas liquid Chromatography</p> <p>Plasmid Fingerprinting</p>	<b>15L</b>

<b>Unit III</b>	<b>Introduction to Immunology</b> 3.1 Basic concepts in immunology Hematopoiesis, Blood cell types, Components of immune system – Cells and organs of the immune system 3.2 Humoral and Cell mediated immune response 3.3 Phagocytosis and Acute and chronic inflammation	<b>15L</b>
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## S.Y.B.Sc. MICROBIOLOGY SYLLABUS

### SEMESTER III MINOR PAPER I

**Course: Environmental Microbiology**

**Course code: SIUMIMN211**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Evaluate and compare methods of air sanitation and quality control.
2. Review microbiology of freshwater, marine, sewage
3. Outline the methods of water potability testing and sewage treatment.
4. Discussing the different soil cycles and evaluating their role in Geomicrobiology

<b>Course code</b>	<b>MINOR PAPER</b>	<b>3 CREDITS</b>
<b>SIUMIMN211</b>	<b>Environmental Microbiology</b>	<b>45 LECTURES</b>
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	<b>Air Microbiology</b> 1.1 Aeromicrobiology: Important airborne pathogens and toxins, Aerosols, nature of bioaerosols, aeromicrobiological pathway, microbial survival in the air, extramural aeromicrobiology, intramural aeromicrobiology 1.2 Sampling Devices for the Collection of Air Samples, Detection of microorganisms on fomites 1.3 Air Sanitation 1.4 Air Quality Standards	<b>15L</b>

<p><b>Unit II</b></p>	<p align="center"><b>Fresh Water and Marine Microbiology</b></p> <p>2.1 Fresh Water Microbiology</p> <ul style="list-style-type: none"> <li>• Fresh water environments and micro-organisms found in Springs, rivers and streams, Lakes,marshes and bogs</li> </ul> <p>2.2 Potable water:</p> <ul style="list-style-type: none"> <li>• Definition, water purification, water quality standards and pathogens transmittedthrough water</li> </ul> <p>2.3 Microbiological analysis of water:</p> <ul style="list-style-type: none"> <li>• Indicator organisms and their detection in water-Total Coliforms, Faecal Coliforms and <i>E. coli</i>, Faecal <i>Streptococci</i>, <i>Clostridium perfringens</i></li> </ul> <p>2.4 Marine Microbiology</p> <ul style="list-style-type: none"> <li>• Marine zones Bacteria in marine, estuarine, ocean environments</li> </ul>	<p align="center"><b>15L</b></p>
<p><b>Unit III</b></p>	<p align="center"><b>Soil and Geo Microbiology</b></p> <p>3.1 Terrestrial Environment</p> <p>Soil- Definition, Composition, function , Textural triangle</p> <p>Types of soil microorganisms and their activities</p> <p>3.2 Methods of studying soil microorganisms:</p> <p>Sampling, Cultural methods, Physiological methods, Immunological methods, Nucleic acidbased methods, Radioisotope techniques</p> <p>3.3 Biogeochemical Cycles:</p> <p>Carbon cycle, Nitrogencycle, Sulphur cycle, Phosphorus Cycle, Iron cycle</p>	<p align="center"><b>15L</b></p>

## S.Y.B.Sc. MICROBIOLOGY SYLLABUS

### SEMESTER III

Course code	PRACTICALS	1 Credit
SIUMIMJP211	MAJOR PAPER I	1 Credit
	TOPIC	
	<ol style="list-style-type: none"><li>1. Estimation of total sugars by Anthrone method Estimation of reducing sugar by DNSA method</li><li>2. Estimation of protein Biuret method (indirect and direct)</li><li>3. Estimation of DNA by DPA method</li><li>4. Estimation of RNA by Orcinol Method</li><li>5. Preparation of Nanoparticles and their characterization using UV Spectrophotometer</li><li>6. Introduction to colorimeter and study of <math>\lambda_{max}</math></li><li>7. Data representation and interpretation</li><li>8. U.V. spectrophotometer (Demo)</li><li>9. DNA estimation by UV Spectrophotometer technique</li><li>10. Working of a pH meter</li></ol>	

Course code	PRACTICALS	1 Credit
SIUMIMJP212	MAJOR PAPER II	1 Credit
	TOPIC	
	<ol style="list-style-type: none"><li>1. Determination of total solids in wastewater</li><li>2. Determination of BOD and COD of wastewater</li><li>3. Visit to a sewage treatment plant or water purification plant</li><li>4. Use of Selective and Differential Solid Media: Mac Conkeys agar, SS agar, XLD agar, TCBS agar, SIBA, Salt Mannitol agar, CLED agar, Cetrinide agar</li><li>5. Blood staining by the Field's method</li><li>6. Preparation of serum and plasma</li></ol>	

<b>Course code</b>	<b>PRACTICALS</b>	<b>1 Credit</b>
<b>SIUMIMNP211</b>	<b>MINOR PAPER I</b>	<b>1 Credit</b>
	<b>TOPIC</b>	
	<ol style="list-style-type: none"> <li>1. Enumeration of microorganisms in air and study of its load after fumigation</li> <li>2. Study of air microflora and determination of sedimentation rate</li> <li>3. Routine analysis of water: Standard Plate Count, Detection of Coliforms in water: Presumptive Test, Confirmed Test , Completed Test</li> <li>4. Study of microbial flora in raw and treated sewage</li> <li>5. Total viable count of soil microflora</li> <li>6. Isolation of bacteria, Actinomycetes and fungi from soil</li> <li>7. Enrichment and isolation of Nitrosifiers, Nitrifiers, Cellulose degraders, Sulphate reducers and Phosphate solubilisers from soil</li> <li>8. Preparation of Winogradsky's column</li> </ol>	

**SEMESTER III  
VOCATIONAL SKILL COURSE  
TECHNIQUES IN MICROBIOLOGY - I**

**Course: VOCATIONAL SKILL COURSE**

**Course code: SIUMIVS211**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Perform lipid extraction using soxhlet apparatus
2. Learn and apply molecular biology technique for the extraction and isolation of DNA from biological sample
3. Estimate DNA quantitatively and visualize by electrophoretic technique
4. Perform water analysis and analyze potability of water
5. Identify pathogens from clinical samples using cultural techniques

<b>Course code</b>	<b>VOCATIONAL SKILL COURSE TECHNIQUES IN MICROBIOLOGY - I</b>	<b>Credits</b>
<b>SIUMIVS211</b>	<b>TOPIC</b>	<b>2 Credits</b>
	<ol style="list-style-type: none"> <li>1. Extraction of lipid by Soxhlet method (Demonstration)</li> <li>2. Isolation and detection of DNA from onion /E.coli</li> <li>3. Demonstration of agarose gel electrophoresis</li> <li>4. Study of biofilm: slide immersion tech and staining</li> <li>5. Routine analysis of water: Standard Plate Count, Detection of Coliforms in water: Presumptive Test, Confirmed Test, Completed Test</li> <li>6. Study of microbial flora in raw and treated sewage</li> <li>7. Use of Biochemical Media/Tests for Identification of Pathogens: Carbohydrate fermentation, Indole test, Methy Red test, Vogues Proskauer test, Citrate Utilization, Lysine Decarboxylase, Gelatin Liquefaction, Nitrate Reduction, Phenylalanine deaminase test, Urease test, TSI agar, Oxidase test, Catalase test, Bile solubility test, Coagulase test, Optochin test and Bacitracin test.</li> </ol>	

## S.Y.B.Sc. MICROBIOLOGY SYLLABUS

### SEMESTER III

#### OPEN ELECTIVE (Interdisciplinary)

**Course: Microbiology in Everyday Life - I**

**Course code: SIUMIOE211**

#### Course Outcomes (CO)

**At the end of the course the students will be able to:**

1. Recognize a prokaryotic and eukaryotic cell
2. List the role of microbes in fermented dairy products
3. Describe the importance of microorganism in human health
4. Define immunity and distinguish between innate and acquired immunity

<b>COURSE CODE</b>	<b>OPEN ELECTIVE</b>	<b>2 CREDIT</b>
<b>SIUMIOE211</b>	<b>Microbiology In Everyday Life - I</b>	<b>15 LECTURES</b>
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	1. Introduction to biology 2. Food microbiology- Fermented foods- Dahi, cheese, butter 3. Industrial production of cheese 4. Microorganisms and human health - Normal flora, Good bacteria, Bad bacteria, Types of Immunity- innate and acquired	15
<b>Unit II</b>	Tutorial 1. Cell types 2. Milk products involving microbes 3. Cheese production 4. Immunity	15

**SYBSc Microbiology Syllabus  
Semester - 4**

**MAJOR PAPER I**

**Course: Metabolism and Basic Analytical Techniques**

**Course code: SIUMIMJ221**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Understanding the principles of bioenergetics with respect to cellular metabolism.
2. Understanding the enzyme kinetics and summarizing the various factors affecting enzyme kinetics.
3. Understanding the principle and working of chromatographic and centrifugation techniques and their applications.

COURSE CODE	MAJOR PAPER I	3 CREDITS
SIUMIMJ221	Metabolism and Basic Analytical Techniques	45 LECTURES 3 CREDITS
UNIT	TOPIC	LECTURES
Unit I	<p><b>Introduction To Metabolism and Bioenergetics</b></p> <p>1.1 Bioenergetics and thermodynamics: Energy transformations, thermodynamic quantities, standard –free energy, difference between <math>\Delta G</math> &amp; <math>\Delta G_0</math></p> <p>1.2 Structure of ATP, phosphoryl group transfer and ATP, Types of energy – rich compounds, multi-roles of ATP, inorganic phosphoryl group donor</p> <p>1.3 Biochemical and chemical reactions, Biological oxidation-reduction reaction</p>	
Unit II	<p style="text-align: center;"><b>Enzyme Kinetics</b></p> <p>2.1 Introduction of Enzymes:</p> <ul style="list-style-type: none"> <li>• General properties of enzymes</li> <li>• Concept of activation energy</li> <li>• Rate law for a simple catalyzed reaction</li> <li>• Michaelis-Menten equation and it's derivation</li> <li>• Lineweaver Burk plot Classification of enzymes</li> <li>• Classification of enzymes</li> </ul>	<b>15L</b>

	<p>2.2 Overview of Coenzyme:</p> <ul style="list-style-type: none"> <li>• Coenzymes: Different types and reactions catalyzed by coenzymes NAD<sup>+</sup>: structure, occurrence and biochemical function</li> </ul> <p>2.3 Enzyme Kinetics:</p> <ul style="list-style-type: none"> <li>• Saturation kinetics</li> <li>• Effect of temperature and pH</li> <li>• Effect of Inhibitors- Reversible and irreversible, competitive, Noncompetitive and uncompetitive inhibitors</li> <li>• Multisubstrate reactions- Ordered, Random and pingpong reactions.</li> <li>• Allosteric effects in enzyme catalysed reactions- Koshland-Nemethy and Filmer model and Monod, Wyman and Changeux model</li> </ul>	
<b>Unit III</b>	<p><b>Analytical techniques</b></p> <p>3.1 Paper and thin layer Chromatography: Introduction to chromatography, types of chromatography Paper chromatography: Principle, circular, ascending and descending Paper Chromatography, Thin layer chromatography :principle, preparation of TLC plates, procedure forTLC, preparative TLC, 2D TLC, HPTLC</p> <p>3.2 Column chromatography : Introduction and principleSize Exclusion chromatography , Ion Exchange chromatography, Affinity chromatography, High Performance Liquid chromatography Gas chromatography</p> <p>3.3 Centrifugation Introduction : basic principles of sedimentation, Types, Preparative centrifugation and its applications, Analytical centrifugation and its applications</p>	<b>15</b>

## MAJOR PAPER II

**Course: Dairy Microbiology, Food Microbiology and Taxonomy**

**Course code: SIUMIMJ222**

### Course Outcomes (CO)

**At the end of the course the students will be able to:**

1. Define the basics of dairy microbiology and applying the role of microbes to develop dairy products.
2. Understand the role of microbes in food microbiology with respect to food production, spoilage and preservation.
3. Classify microorganisms based on taxonomic principles and evaluate the different methods of microbial taxonomy.

<b>COURSE CODE</b>	<b>MAJOR PAPER II</b>	<b>3 CREDITS 45 LECTURES</b>
<b>SIUMIMJ222</b>	<b>Dairy Microbiology, Food Microbiology and Taxonomy</b>	
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	<b>Dairy Microbiology</b> 1.1 Milk- Definition, composition, Spoilage of milk: Sources of contamination of milk. Color defects, Flavour defects, Ropiness of milk Stormy fermentation of milk 1.2 Pasteurization of milk <ul style="list-style-type: none"><li>• LTLT, HTST, UHT Method</li><li>• Efficiency testing of Pasteurization (Phosphatase Test)</li><li>• Milk products- production of a Yoghurt, Butter, Cheese-Cheddar and Cottage cheese, Dried milk.</li></ul> 1.3 Quality control of milk:- <ul style="list-style-type: none"><li>• Rapid platform test:- MBRT, Resazurin</li><li>• Microbiological analysis of milk:- SPC, Coliformcount, Psychrophiles, Thermophilic count.</li></ul>	<b>15L</b>

<p><b>Unit II</b></p>	<p><b>Food Microbiology</b></p> <p>2.1 Introduction: Significance, food as a substrate and sources, Intrinsic and extrinsic factors</p> <p>2.2 General Principles of spoilage: Spoilage of fresh foods: fruits and vegetables, eggs, meat, poultry and seafood.</p> <p>2.3 General principles of food preservation (principle of each method and example of foods only): High temperature (Including TDT, TDP, D, F and Z value), Low temperature (Freezing), Asepsis, Drying, Radiations, Chemical preservatives</p> <p>2.4 Food control agencies:- HACCP, FDA, FSSAI</p> <p>2.5 Methods of detection of microorganisms in food: overview of cultural, microscopic, physical, chemical and bioassay methods</p>	<p><b>15L</b></p>
<p><b>Unit III</b></p>	<p><b>Microbial Taxonomy</b></p> <p>3.1 Introduction to microbial taxonomy</p> <p>3.2 Systems of classification (Cavalier Smith 6 kingdom) Bergey's manual</p> <p>3.3 The three domain concept based on phylogeny- Nomenclature, Taxonomic ranks, Numerical Taxonomy</p> <p>3.4 Methods of analysis used in classification: Phenotypic analysis (Morphological characteristics Physiological and metabolic characteristics, Biochemical characteristics, Ecological characteristics, Fatty acid analysis).</p> <p>3.5 Microarray Nucleic acid sequencing types, Phylogenetic tree: concept</p>	<p><b>15</b></p>

**SYBSc Microbiology Syllabus  
Semester 4**

**Course: Industrial Microbiology, Microbial Diversity and Applications of Microbiology**

**Course code: SIUMIMN221**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Describe and understand basic fundamentals of industrial microbiology.
2. Classifying microorganisms based on taxonomic principles and evaluate the different methods of microbial taxonomy.
3. Understanding extreme environment and comparing the microbial diversity and studying the applications of extremophiles.
4. Evaluating the role of microbes as biofertilizers, biocontrol agents and remediation of polluted environment.

<b>COURSE CODE</b>	<b>MINOR PAPER</b>	<b>3 CREDITS</b>
<b>SIUMIMN221</b>	<b>Industrial Microbiology, Microbial Diversity and Applications of Microbiology</b>	<b>45 LECTURES</b>
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	<b>Industrial Microbiology</b> 1.1 Strains of industrially important microorganisms: Desirable characteristics of industrial strain Principles and methods of primary and secondary Screening 1.2 Types of fermentations: Surface and Submerged, Batch, continuous fermentation Solid state fermentation 1.3 Design of an ideal fermenter: conventional stirred tank reactor 1.4 Concept of upstream and Downstream processing Industrial inoculum build up	<b>15L</b>

	<p>1.5 Media for industrial fermentations:</p> <p>Production and Inoculum media,</p> <p>Media components :- Carbon source, nitrogen source, amino acids and vitamins, minerals, water, buffers, antifoam agents, precursors, inhibitors and inducers</p>	
<b>Unit II</b>	<p><b>Biodiversity in extreme environments and Metagenomics</b></p> <p>2.1 Biodiversity in extreme environments</p> <ul style="list-style-type: none"> <li>• Extreme Environments and their types with respect to the physical conditions which lead to microbial stress</li> <li>• Temperature based environments- Low and high temperature environments</li> <li>• pH based environments- Acidic and alkaline environments, Acid mine drainage</li> <li>• Environments with high salt concentration</li> </ul> <p>2.2 Applications of extremophiles:</p> <ul style="list-style-type: none"> <li>• Applications of Acidophiles and Alkalophiles</li> <li>• Applications of halophiles- in biotechnology and medicine</li> <li>• Applications of psychrophiles in pharmaceuticals and environment.</li> <li>• Applications of thermophiles and hyperthermophiles in enzymology</li> </ul>	<b>15L</b>

<b>Unit III</b>	<p align="center"><b>Biofertiliser, Biopesticide , Bioremediation</b></p> <p>3.1 Biofertiliser: Introduction of Biofertilizers, Different types of biofertilizers, Mass production of Biofertilizers, Application of Biofertilizers</p> <p>3.2 Biopesticides- Introduction of biopesticides, Types of Biopesticides Technical Aspects of Biopesticides, Major biopesticides produced and used in India</p> <p>3.3 Bioremediation: Introduction of Bioremediation, Principle of Bioremediation Factors affecting Bioremediation, Phytoremediation – types, Microbial Populations used for Bioremediation processes, Bioremediation strategies, Advantages and Disadvantages of Bioremediation</p>	<b>15</b>
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**SYBSc Microbiology Syllabus  
Semester 4**

<b>COURSE CODE</b>	<b>PRACTICALS</b>	<b>1 Credits</b>
<b>SIUMIMJP221</b>	<b>MAJOR PAPER I</b>	<b>1 Credit</b>
	<ol style="list-style-type: none"> <li>1. Problems on bioenergetics to calculate the <math>K_{eq}</math>; Gibbs energy, enthalpy, etc</li> <li>2. Effect of pH, Temp, substrate and enzyme concentration on activity of invertase.</li> <li>3. Determination of <math>K_m</math> and <math>V_{max}</math> of an enzyme</li> <li>4. Separation and identification of amino acids by ascending paper chromatography</li> <li>5. Density gradient centrifugation and sizing of yeast cells</li> </ol>	

<b>COURSE CODE</b>	<b>PRACTICALS</b>	<b>1 Credits</b>
<b>SIUMIMJP222</b>	<b>MAJOR PAPER II</b>	<b>1 Credit</b>
	<ol style="list-style-type: none"> <li>1. Visit to Food/Dairy industry</li> <li>2. RPT of Milk– RRT, MBRT, DMC</li> <li>3. Isolation of food spoilage agent:</li> <li>4. Fruit/Vegetable- Physical and Microscopic and Pectinolytic agent</li> <li>5. Meat - Proteolytic, lipolytic, saccharolytic</li> <li>6. Identification of bacteria using Bergey's Manual</li> </ol>	

<b>COURSE CODE</b>	<b>PRACTICALS</b>	<b>1 Credits</b>
<b>SIUMIMNP221</b>	<b>MINOR PAPER</b>	<b>1 Credit</b>
	<b>TOPIC</b>	
	<ol style="list-style-type: none"> <li>1. Crowded plate technique</li> <li>2. Antibiotic producer screening by Wilkins overlay method</li> <li>3. Antibiotic spectrum determination(streak/strip)</li> <li>4. Solid state fermentations (group experiment)</li> <li>5. Isolation of thermophiles, acidophiles and psychrophiles</li> <li>6. Estimation of heavy metals (Titration method)</li> <li>7. Preparation of biofertilizer</li> <li>8. Efficacy of biofertilizer</li> </ol>	

**SEMESTER IV**

**SKILL ENHANCEMENT COURSE**

**TECHNIQUES IN MICROBIOLOGY - II**

**Course: SKILL ENHANCEMENT COURSE**

**Course code: SIUMISE221**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Perform primary screening of potential industrially important enzyme producing microbes
2. Analyze quality of milk as per BIS and FSSAI standards
3. Comprehend physical and chemical methods of food preservation.

<b>COURSE CODE</b>	<b>PRACTICALS</b>	<b>1 Credits</b>
<b>SIUMISE221</b>	<b>SKILL ENHANCEMENT COURSE TECHNIQUES IN MICROBIOLOGY - II</b>	<b>1 Credit</b>
	<b>TOPIC</b>	
	<ol style="list-style-type: none"><li>1. Isolation of amylase, protease, lipase producers</li><li>2. Extracellular production of invertase from yeast</li><li>3. Microbiological Quality Control of Milk as per BIS/FSSSAI</li><li>4. Analysis of Cheese, Paneer, Butter, Yogurt/curd as per BIS/FSSAI (Group experiment)</li><li>5. Determination of TDT and TDP</li><li>6. Determination of Salt and sugar tolerance</li><li>7. Determination of MIC of a Chemical preservative</li></ol>	

**SYBSc Microbiology Syllabus  
Semester 4  
OPEN ELECTIVE (Interdisciplinary)**

**Course: Microbiology in Everyday Life - II**

**Course code: SIUMIOE221**

**Course Outcomes (CO)**

**At the end of the course the students will be able to:**

1. Recognize a prokaryotic and eukaryotic cell
2. Outline the microbes that cause common infections like Malaria, dengue, typhoid, COVID
3. List the role of microbes in agriculture
4. Review the use of microorganisms for antibiotic production
5. Define antimicrobial drug resistance

<b>COURSE CODE</b>	<b>OPEN ELECTIVE</b>	<b>2 CREDIT</b>
<b>SIUMIOE221</b>	<b>Microbiology In Everyday Life - II</b>	<b>15 LECTURES 1 CREDITS</b>
<b>UNIT</b>	<b>TOPIC</b>	<b>LECTURES</b>
<b>Unit I</b>	1. Introduction to biology 2. Microbes and agriculture -Biofertilizers, biopesticides, bioherbicides 3. Microorganisms and human health -Malaria, dengue, typhoid, COVID 4. Antibiotic production and drug resistance	15
<b>Unit II</b>	Tutorial 1. Prokaryotic cell & Eukaryotic cell 2. Microbes and agriculture 3. Vector borne diseases 4. Antibiotic production and drug resistance	15

### References for MAJOR Papers

1. Fundamentals of Biochemistry. D. Voet and J. Voet Publisher Wiley plus Edition 5th.
2. Instrumental Methods of chemical analysis, V.K. Ahluwalia, Ane Books Pvt.Ltd; 2015.
3. Laboratory manual in Biochemistry- J.Jayaraman.
4. Lehninger- Principles of Biochemistry- David Nelson, Michael Cox. 4th edition W.H. Freeman and Company [Low price edition- for sale in India, Pakistan, Sri Lanka, Bangladesh, Nepal and Bhutan
5. Bionanotechnology - Andrew and Waqar, One Central Press Ltd, UK., November, 2014.
6. Brock biology of microorganism by Michael T Madigan. and John M
7. Genetics-A molecular approach, Peter J. Russell (2006) 2nd ed.
8. Diagnostic Microbiology, Bailey and Scott, 11th edition Publ: Mosby
9. Immunology Essential and Fundamental, Third Edition, Pathak and Palan.
10. Immunology; Kuby 6th edition.
11. Microbiology By Prescott, Harley, Klein's 7th Edn
12. Microbiology, An Introduction by Tortora, Funke and Case 9<sup>th</sup> and 11th edition, Pearson education.
13. Understanding Enzymes: Trevor Palmer 4<sup>th</sup> Edition.

### References for Minor Paper

1. Air Quality Standards- NAAQS Manual , Volume I
2. Environmental Microbiology , 2nd Edition; Raina M. Maier, Ian L. Pepper, Charles P. Gerba, 2010 Academic Press
3. Fundamental Principles of Bacteriology , 7th Edition; A.J. Salle , Tata Mc Graw Hill Publishing Company
4. Fundamentals of Microbiology, 9th Edition , Frobisher, Hinsdill, Crabtree, Goodheart, 1974, Saunders College Publishing
5. Introduction to Environmental Microbiology – Barbara Kolwzan, Waldemar Adamiak.
6. Prescott's Microbiology, 8th Edition; Joanne M. Willey, Linda M. Sherwood, Christopher J. Woolverton, 2011, Mc Graw Hill International Edition
7. Soil Microbiology-4th Edition, N.S Subba Rao, 2000, Oxford and IBH Publishing Pvt Ltd  
Biopesticides: An eco-friendly approach for pest control; Journal of Biopesticides 3(1 Special Issue) 186 - 188 (2010) 186, Suman Gupta and A. K. Dikshit
8. Bioremediation - An Overview Jr. of Industrial Pollution Control 27(2)(2011) pp 161-168, V. Mary Kensa
9. Bioremediation: Features, Strategies and applications, Shilpi Sharma.
10. Brock Biology of Microorganisms, Madigan, Martinko, Dunlap and Clark (2009) 12<sup>th</sup> edition, Pearson Education

### References for VSC & SEC

1. A handbook book of Organic analysis: qualitative and quantitative 4th edition, Hans Thacher Clarke, CBS publishers and distributors, New Delhi.
2. An Introduction To Practical Biochemistry by Plummer David (1979)TMH
3. Laboratory Manual in Biochemistry, J. Jayaraman, (2003) New Age International Publishers
4. Methods In Microbiology, Vol.5B, Ed. Norris and Ribbon, Academic Press
5. Principles and Techniques of practical biochemistry by William and Wilson.
6. Principles and techniques of Biochemistry and Molbiology 6<sup>th</sup> ed, Keith Wilson and John Walker, Cambridge University press,2006
7. Principles of Biochemistry, Lehninger: 4<sup>th</sup> Ed.,D.Nelson and M. Cox, W. H. Freeman and Co., (LPE)
8. Practical medical microbiology by Mackie and McCartney 14th edition. Publ:Churchill Livingstone
9. Textbook of Microbiology by Anantnarayan and Paniker's, 8<sup>th</sup> Ed.
10. The Elements of Immunology by Fahim Khan. Pearson Education.